

## Follow the general instructions below to prepare for bone marrow sample collection.

### Pre-Collection:

1. Clearly identify the patient.
2. Label slide, tubes, and containers with 2 patient identifiers.
  - 2.1. Acceptable patient identifiers: patient's full name, date of birth, medical record number.
3. Assemble collection material and tubes.
4. Standard bone marrow collection:
  - 4.1. Initial non-heparinized syringe: 0.5 to 1 mL of bone marrow aspirate
  - 4.2. 2 lavender top (EDTA) tubes: 2-3mL each for molecular testing
  - 4.3. 2 green top (sodium heparin) tubes: 3mL each for flow cytometry/chromosome/FISH studies
  - 4.4. 1-2 containers filled with B-plus fixative
  - 4.5. 1 conical tube filled with RPMI for fresh biopsy if dry tap (optional at hospital sites).
  - 4.6. 1 non-additive red top tube: 0.5 mL of non-heparinized aspirate for clot
  - 4.7. 10 clean glass slides on work surface for collection. Have extra slides available for use if needed.
    - 4.7.1. ≈7 slides (bone marrow aspirate smears) and ≈3 slides (biopsy touch preps)
5. MRD (minimal residual disease testing):
  - 5.1. Collect an additional 1-2 mL of bone marrow aspirate as the first aspirate in sodium heparin or EDTA tube
  - 5.2. Continue collection as stated in 3.
6. Engraftment (chimerism) studies:
  - 6.1. Collect standard bone marrow collection and
  - 6.2. 2 mL of additional bone marrow aspirate in lavender top (EDTA) per cell type ordered.
7. Paperwork/tubes/containers must include biopsy site location (i.e. right iliac crest) & collection time.

### Peripheral blood smears made from venipuncture specimen:

1. Collect 3mL peripheral blood sample via venipuncture into an EDTA.
2. Make 2 direct smears manually, adjusting as necessary for proper length and thickness.
3. Smears can be made by the processing site if the sample arrives within 16 hours of collection.

### Collection: Follow steps below to obtain bone marrow core biopsy, clot, and aspirate specimens:

1. Syringes used for bone marrow slides, MRD testing, clot should not be rinsed with heparin.
  - 1.1. **Do not exceed 2mL** as this dilutes specimen and reduces quality of the specimen.
2. All other syringes can be pre-rinsed with liquid heparin to prevent clotting.
3. Expel some of the non-heparinized aspirate on a slide and check for spicules.
  - 3.1. If adequate spicules are present, continue.
  - 3.2. If the sample is inadequate, request redirect of needle for better spicule sample.
  - 3.3. Due to therapy or disease, some samples may not have good spicules.
    - 3.3.1. An additional core biopsy can be placed in RPMI for additional studies when aspirate is inadequate
4. Make slides immediately for the aspirate collected using non-heparinized syringe.
  - 4.1. Tip the slide so excess fluid drains away from spicules.
  - 4.2. Prepare slides evenly without crush artifact, of correct length and thickness.
  - 4.3. If slides are not prepared during collection, place aspirate syringe contents into EDTA tube.

# Geisinger BONE MARROW REQUISITION

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Requesting Location _____ Location Code _____ Requesting Provider _____		Coll. Date: _____ Rcv Date: _____ Coll. Time: _____ Rec Time: _____ Tech Code: _____		<b>Special Instructions:</b> All specimens to arrive within stability limits or special courier needed. _____ _____	
<b>Diagnosis Code(s)</b> 1. _____ 2. _____ 3. _____ 3. _____					
<b>PATIENT HISTORY</b> _____ _____ History of Bone Marrow Transplant: _____ No _____ Allogenic _____ Auto _____ History of Chemotherapy: _____ No _____ Medications _____					
<b>All Specimens and Slides must be labelled with 2 identifiers</b> Document # specimens Peripheral Blood (PB) ___ Bone Marrow (BM) EDTA ___ Heparin ___ Biopsy ___ Clot Document # slides PB _____ BM _____					
<b>SOURCE: BLOOD</b>			<b>REQUIRED SPECIMEN</b>		
<input type="checkbox"/> PERIPHERAL BLOOD SMEAR			<ul style="list-style-type: none"> <li>• CBC and Diff results/OR lavender top EDTA tube (refrigerated), 16-hour stability.</li> <li>• Slide (stained or unstained).</li> <li>• If previously performed in 16 hours, no additional tube needed.</li> </ul>		
<b>SOURCE: BONE MARROW</b>			<b>SUBMIT IN BONE MARROW TRANSPORT KIT (ROOM TEMP):</b>		
<input type="checkbox"/> BONE MARROW Site _____ B+ fixation			<ul style="list-style-type: none"> <li>• Bone Marrow Core Needle Biopsy in B-Plus fixative</li> </ul>		
<input type="checkbox"/> RPMI COLLECTION (FOR DRY TAP BONE MARROWS)+++			<ul style="list-style-type: none"> <li>• Bone Marrow Aspirate in RPMI solution</li> </ul>		
<input type="checkbox"/> BONE MARROW ASPIRATE SLIDES			<ul style="list-style-type: none"> <li>• 7 Unfixed / unstained / air dried bone marrow slides (2 Push 2 Squash, and 3 Extras)</li> </ul>		
<input type="checkbox"/> BONE MARROW BIOPSY TOUCH IMPRINT			<ul style="list-style-type: none"> <li>• 3 Unfixed / unstained / air dried bone marrow biopsy touch slides</li> <li>• Any remaining aspirate in a lavender top (indicate "bone marrow" on tube)</li> </ul>		
<input type="checkbox"/> BONE MARROW ASPIRATE CLOT			<ul style="list-style-type: none"> <li>• Bone Marrow Aspirate in non-additive container</li> </ul>		
<input type="checkbox"/> ANCILLARY TESTING CYTOGENETICS (CHROMOSOME ANALYSIS) FLOW CYTOMETRY +++ MOLECULAR DIAGNOSTICS +++			<ul style="list-style-type: none"> <li>• 2 sodium heparin tubes with 3 mL each (minimum) of bone marrow</li> <li>• 2 EDTA tubes with 2 mL each (minimum) of bone marrow</li> </ul>		
<input type="checkbox"/> MINIMUM RESIDUAL DISEASE ___ ALL ___ AML ___ NPM1			<ul style="list-style-type: none"> <li>• First 1-2 mL non-heparinized tube of bone marrow aspirate</li> </ul>		
<input type="checkbox"/> ENGRAFTMENT STUDIES (CHIMERISM) ___ T CELL ___ NK CELL ___ MYELOID CELL			<ul style="list-style-type: none"> <li>• 2 mL of additional bone marrow aspirate in lavender top (EDTA) tube per cell type ordered</li> </ul>		
<input type="checkbox"/> _____ B CELL _____ MONOCYTE					